



## 16<sup>th</sup> INTERNATIONAL CONFERENCE ON CARBON DIOXIDE UTILIZATION

### Continuous Production of 2,3-butanediol from CO<sub>2</sub> by *Cupriavidus necator* gas fermentation

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The world is still producing industrial chemicals from finite resources and emit CO<sub>2</sub> as by-product. To sustain our chemical production and avoid catastrophic climate change, alternative ways of producing chemicals must be implemented. Microbial gas fermentation of CO<sub>2</sub> has the potential of addressing the sustainability challenge of chemical industry. The process uses renewable raw material and is carbon neutral or negative [1]. The rapid advancement of synthetic biology provides tools and strategies to modify the host gas fermentation bacteria for producing various chemicals. This study demonstrates the production of 2,3-butanediol (2,3-BDO), a precursor to plastics and pesticides [2], using gas fermentation of *Cupriavidus necator* from CO<sub>2</sub>.

*C. necator*, capable of chemolithotrophic growth on CO<sub>2</sub>, H<sub>2</sub> and air, was modified to host the 2,3-BDO production pathway. The pathways, containing genes: alsS (acetolactate synthase), alsD (acetolactate decarboxylase) from *Bacillus subtilis* and either NADH-dependent BDH (butanediol dehydrogenase) or NADPH dependent sADH (secondary alcohol dehydrogenase) from clostridium, were constructed as an operon under arabinose inducible promoter and integrated to the chromosome.

production strains at lab scale. The main features of the process are presented in Figure 1. In this design, H<sub>2</sub> was introduced to the bioreactor through a separate sparger. The headspace O<sub>2</sub> concentration was monitored and controlled at the set point of 4% (below the O<sub>2</sub> percentage corresponding to the upper explosive limit of H<sub>2</sub>). A PID controller was set up to regulate the air flow rate by the headspace O<sub>2</sub> concentration. This improved the oxygen transfer rate, and allowed high cell density (maximum nearly 50 g dry cell weight/L) at steady state. 2,3-BDO concentration in the two different strain variants during the steady state was 7 g/L, corresponding to a productivity of 0.35 g/L<sup>-1</sup>h<sup>-1</sup>. This is the highest productivity of 2,3-BDO reported to date in autotrophic cultures on CO<sub>2</sub> of *C. necator*. And the process is still being optimized.

The results suggest the biochemical production of 2,3-BDO can be further optimized to a viable industrial process. More importantly, as a successful case study, producing chemicals from CO<sub>2</sub> through gas fermentation was demonstrated to be a promising alternative to conventional petrochemical processes. The understanding and ability to manipulate biological metabolism will continue to expand the product portfolio and market of chemicals produced from CO<sub>2</sub>.

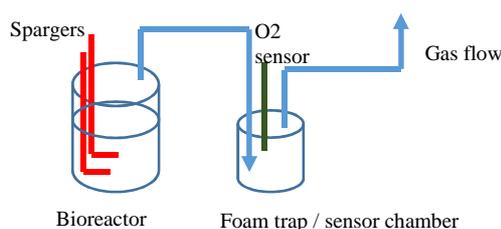


Figure 1. Schematic diagram of the gas fermentation process.

A phosphorous limiting chemostat gas fermentation process was developed to characterize the 2,3-BDO

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#### References

- [1] Md. S. Islam Mozumder, L. Garcia-Gonzalez, H. De Wever, E.I.P. Volcke, *Biochem. Eng. J* **2015**, 98, 107.
- [2] X. Ji, H. Huang, P. Ouyang, *Biotechnology Advances* **2011**, 29, 351.